(FILE 'HOME' ENTERED AT 14:06:56 ON 11 MAR 2004)

INDEX 'ADISCTI, ADISINSIGHT, ADISNEWS, AGRICOLA, ANABSTR, AQUASCI, BIOBUSINESS, BIOCOMMERCE, BIOSIS, BIOTECHABS, BIOTECHDS, BIOTECHNO, CABA, CANCERLIT, CAPLUS, CEABA-VTB, CEN, CIN, CONFSCI, CROPB, CROPU, DISSABS, DDFB, DDFU, DGENE, DRUGB, DRUGMONOG2, ...' ENTERED AT 14:07:12 ON 11 MAR 2004

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SEA (PSYCHROTROPH? OR PSYCHROPHIL? OR FLUORESCENS? OR SYRINGAE?
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35 FILE AOUASCI
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FILE 'SCISEARCH, CABA, CAPLUS, BIOTECHNO, LIFESCI, ESBIOBASE, GENBANK, DGENE, BIOSIS, EMBASE, USPATFULL, PASCAL, BIOTECHDS, AGRICOLA, MEDLINE'

QUE (PSYCHROTROPH? OR PSYCHROPHIL? OR FLUORESCENS? OR SYRINGAE?

- ENTERED AT 14:11:33 ON 11 MAR 2004 L2 9026 S (PSYCHROTROPH? OR PSYCHROPHIL? OR FLUORESCENS? OR SYRINGAE?)
- L3 2256 S L2 (S)(ISOLAT? OR PURIF?)
- L4 1886 S L3 (S) (VECTOR? OR GENE? OR POLYNUCLEOT? OR INSERT? OR DNA?
- L5 732 S L4 (S) FLUORESCENS?
- L6 368 S L5 (S) COLI

Ll

L7 161 DUP REM L6 (207 DUPLICATES REMOVED)

- L8 L9
- 103 S L7 AND PY<=1999 259 S L4 (S) (HEAT? OR TEMPERAT?) 133 DUP REM L9 (126 DUPLICATES REMOVED) 133 SORT L10 PY A L10
- LII

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PASSWORD:

TERMINAL (ENTER 1, 2, 3, OR ?):2

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                 "Ask CAS" for self-help around the clock
NEWS
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         SEP 09
                 CA/CAplus records now contain indexing from 1907 to the
                 present
NEWS
         DEC 08
                 INPADOC: Legal Status data reloaded
         SEP 29
NEWS
      5
                 DISSABS now available on STN
NEWS
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                 BIOTECHNO no longer updated
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                 German (DE) application and patent publication number format
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              AND CURRENT DISCOVER FILE IS DATED 3 MARCH 2004
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              CAS World Wide Web Site (general information)
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Enter NEWS followed by the item number or name to see news on that specific topic.

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FILE 'HOME' ENTERED AT 14:06:56 ON 11 MAR 2004

=> index bioscience medicine
FILE 'DRUGMONOG' ACCESS NOT AUTHORIZED
COST IN U.S. DOLLARS

72

FILE WPINDEX FILE NLDB

INDEX 'ADISCTI, ADISINSIGHT, ADISNEWS, AGRICOLA, ANABSTR, AQUASCI, BIOBUSINESS, BIOCOMMERCE, BIOSIS, BIOTECHABS, BIOTECHDS, BIOTECHNO, CABA, CANCERLIT, CAPLUS, CEABA-VTB, CEN, CIN, CONFSCI, CROPB, CROPU, DISSABS, DDFB, DDFU, DGENE, DRUGB, DRUGMONOG2, ... 'ENTERED AT 14:07:12 ON 11 MAR 2004

71 FILES IN THE FILE LIST IN STNINDEX

Enter SET DETAIL ON to see search term postings or to view search error messages that display as 0* with SET DETAIL OFF.

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=> s (psychrotroph? or psychrophil? or fluorescens? or syringae?) (s) (express?)
              FILE ADISNEWS
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         2
              FILE ANABSTR
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              FILE AQUASCI
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              FILE BIOBUSINESS
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       687
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             FILE NTIS
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48 FILES HAVE ONE OR MORE ANSWERS, 71 FILES SEARCHED IN STNINDEX

L1 QUE (PSYCHROTROPH? OR PSYCHROPHIL? OR FLUORESCENS? OR SYRINGAE?) (S) (EXPR ESS?)

=> d rankn DISPLAY L# IS NOT VALID IN STNINDEX Answer set was created in a file. Enter DISPLAY HISTORY to see where the answer set was created. Use the File command to change to that file, then display the answer.

. d wante		
=> d rank	927	SCISEARCH
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F35	8	DRUGU
F36	7	EMBAL
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F38	5	DDFU
F39	5	NTIS
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F44	2	CONFSCI
F45	2	VETU
F46	1	ADISNEWS
F47	1	CEN
F48	1	HEALSAFE
110	*	THE THOUSE IS

=> file f1-f16 COST IN U.S. DOLLARS

SINCE FILE TOTAL ENTRY SESSION 3.99 4.20

FULL ESTIMATED COST

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FILE 'MEDLINE' ENTERED AT 14:11:33 ON 11 MAR 2004
=> s (psychrotroph? or psychrophil? or fluorescens? or syringae?) (s) (express?)
  8 FILES SEARCHED...
        9026 (PSYCHROTROPH? OR PSYCHROPHIL? OR FLUORESCENS? OR SYRINGAE?)
               (S) (EXPRESS?)
=> s l2 (s)(isolat? or purif?)
  8 FILES SEARCHED...
         2256 L2 (S) (ISOLAT? OR PURIF?)
=> s 13 (s) (vector? or gene? or polynucleot? or insert?)
  4 FILES SEARCHED...
  6 FILES SEARCHED...
  7 FILES SEARCHED...
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SEARCH ENDED BY USER
  8 FILES SEARCHED...
SEARCH ENDED BY USER
=> s 13 (s) (vector? or gene? or polynucleot? or insert? or dna? or plasmid?)
  4 FILES SEARCHED...
  6 FILES SEARCHED...
  7 FILES SEARCHED...
  8 FILES SEARCHED...
 10 FILES SEARCHED...
 12 FILES SEARCHED..
         1886 L3 (S) (VECTOR? OR GENE? OR POLYNUCLEOT? OR INSERT? OR DNA? OR
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PLASMID?)

=> s 14 (s) fluorescens?

isolate.

- L11 ANSWER 105 OF 133 USPATFULL on STN
- TI Recombinant bacterial phytases and uses thereof
- L11 ANSWER 106 OF 133 USPATFULL on STN
- TI Novel antigen binding molecules for therapeutic, diagnostic, prophylactic, enzymatic, industrial, and agricultural applications, and methods for generating and screening thereof
- L11 ANSWER 107 OF 133 USPATFULL on STN
- TI Non-stochastic generation of genetic vaccines
- L11 ANSWER 108 OF 133 USPATFULL on STN
- TI End selection in directed evolution
- L11 ANSWER 109 OF 133 USPATFULL on STN
- TI Receptors for hypersensitive response elicitors and uses thereof
- L11 ANSWER 110 OF 133 USPATFULL on STN
- TI Saturation mutagenesis in directed evolution
- L11 ANSWER 111 OF 133 USPATFULL on STN
- TI Enzymes having alpha amylase activity and methods of use thereof
- L11 ANSWER 112 OF 133 USPATFULL on STN
- TI Synthetic ligation reassembly in directed evolution
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- TI Enzymes having alpha amylase activity and methods of use thereof
- L11 ANSWER 114 OF 133 USPATFULL on STN
- TI Recombinant constructs and systems for secretion of proteins via type III secretion systems
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- TI Enzymes having alpha amylase activity and methods of use thereof
- L11 ANSWER 116 OF 133 USPATFULL on STN
- TI Phytases, nucleic acids encoding them and methods for making and using them
- L11 ANSWER 117 OF 133 USPATFULL on STN
- TI Recombinant phytases and uses thereof
- L11 ANSWER 118 OF 133 USPATFULL on STN
- TI Saturation mutagenesis in directed evolution
- L11 ANSWER 119 OF 133 USPATFULL on STN
- TI Enzymes having glycosidase activity and methods of use thereof
- L11 ANSWER 120 OF 133 USPATFULL on STN
- TI Synthetic ligation reassembly in directed evolution
- L11 ANSWER 121 OF 133 USPATFULL on STN
- TI Recombinant bacterial phytases and uses thereof
- L11 ANSWER 122 OF 133 USPATFULL on STN
- TI Enzymes having carboxymethyl cellulase activity and methods of use thereof
- L11 ANSWER 123 OF 133 USPATFULL on STN
- TI Exonuclease-mediated nucleic acid reassembly in directed evolution
- L11 ANSWER 124 OF 133 USPATFULL on STN
- TI Novel methods of enzyme purification
- L11 ANSWER 125 OF 133 BIOTECHDS COPYRIGHT 2004 THOMSON DERWENT/ISI on STN
- TI Recombinant cold-adapted trypsin I from Atlantic cod-expression, purification, and identification;
 - recombinant enzyme production via plasmid expression in host cell for

use in medicine and flavor

L11 ANSWER 126 OF 133 SCISEARCH COPYRIGHT 2004 THOMSON ISI on STN
TI Cold-active esterase from Psychrobacter sp Ant300: gene cloning,
characterization, and the effects of Gly -> Pro substitution near the
active site on its catalytic activity and stability

L11 ANSWER 127 OF 133 SCISEARCH COPYRIGHT 2004 THOMSON ISI on STN Recombinant cold-adapted trypsin I from Atlantic cod-expression, purification, and identification

L11 ANSWER 128 OF 133 USPATFULL on STN
TI End selection in directed evolution

L11 ANSWER 129 OF 133 USPATFULL on STN

TI Identification of essential genes in microorganisms

L11 ANSWER 130 OF 133 USPATFULL on STN
TI End selection in directed evolution

L11 ANSWER 131 OF 133 USPATFULL on STN

TI Enzymes having secondary amidases activity and methods of use thereof

L11 ANSWER 132 OF 133 USPATFULL on STN

TI Phospholipases, nucleic acids encoding them and methods for making and using them

L11 ANSWER 133 OF 133 USPATFULL on STN

TI Synthetic ligation reassembly in directed evolution

=> d 111 ibib abs 2 8 11 18 26 28 30 34 49 51 65 67 73 99 100 124

L11 ANSWER 2 OF 133 BIOTECHDS COPYRIGHT 2004 THOMSON DERWENT/ISI on STN

ACCESSION NUMBER: 2002-10994 BIOTECHDS

TITLE: Identifying bioactivities or biomolecules by screening clones

from a gene library generated from more than one organism; enzyme identification using high throughput screening of Streptomyces venezuelae, Escherichia coli, Actinomyces sp.

DNA library

AUTHOR: SHORT J M; KELLER M

PATENT ASSIGNEE: DIVERSA CORP

PATENT INFO: US 2002001809 3 Jan 2002 APPLICATION INFO: US 1997-848095 16 Jun 1997 PRIORITY INFO: US 2001-848095 3 May 2001

DOCUMENT TYPE: Patent LANGUAGE: English

OTHER SOURCE: WPI: 2002-194904 [25]

AN 2002-10994 BIOTECHDS

AB DERWENT ABSTRACT:

NOVELTY - A method for identifying bioactivities or biomolecules, comprising inserting a bioactive substrate into clones from a gene library generated from more than one organism and screening the clones for a change in the substrate, is new.

DETAILED DESCRIPTION - A method for identifying bioactivities or biomolecules using high-throughput screening of nucleic acids comprising: (a) providing a gene library comprising several clones (the nucleic acid for generating the library is obtained from more than one organism); (b) inserting a bioactive substrate into the clones (a bioactivity or biomolecule produced by the clones is detectable by a difference in the substrate before and after contact with the clones); (c) screening the clones with an assay or analyzer that detects a bioactivity or biomolecule; and (d) identifying clones detected as positive for a change in the substrate (a change in the substrate is indicative of DNA that encodes a bioactivity or biomolecule).

BIOTECHNOLOGY - Preferred Method: The clones and substrate are encapsulated in gel microdroplets before screening, optionally together with an indicator cell. The samples are **heated** before step (b), preferably at 70degreesC for 30 minutes. The bioactive substrate is 5-dodecanoylamino-fluorescein-di-D-galactopyranoside (C12FDG) or another

compound with a lipophilic tail. The library is biopanned and/or normalized before step (b). The microdroplets are screened using a fluorescence analyzer, especially a fluorescence-activated cell sorting (FACS) apparatus, or a chromogenic analyzer or by immunoassay. Preferred Library: The gene library is an expression library generated from extremophile DNA in prokaryotic cells, either directly in Streptomyces cells, especially Streptomyces venezuelae, or in Escherichia coli cells followed by transfer to a myceliate bacterium or fungus, preferably an Actinomyces or Streptomyces species, especially Streptomyces venezuelae.

USE - The method is especially useful for identifying enzymes in extremophiles, especially where the enzymes are lipases, esterases, proteases, glycosidases, glycosyl transferases, phosphatases, kinases, mono- and dioxygenases, haloperoxidases, lignin peroxidases, diarylpropane peroxidases, epoxide hydrolases, nitrile hydratases, nitrilases, transaminases, amidases or acylases, and the extremophiles are thermophiles, hyperthermophiles, psychrophiles, halophiles, psychrotrophs, alkalophiles or acidophiles.

ADVANTAGE - The method can be applied to nucleic acids isolated directly or indirectly from the environment using flow cytometry systems normally used for sorting eukaryotic cells.

EXAMPLE - No relevant examples are given. (40 pages)

L11 ANSWER 8 OF 133 BIOTECHDS COPYRIGHT 2004 THOMSON DERWENT/ISI on STN

ACCESSION NUMBER: 2003-14535 BIOTECHDS

TITLE: Detecting hydrolase activity, useful particularly for

identifying variant enzymes with altered properties,

comprises detecting acetic acid released from acetate ester;

stereospecific substrate for Pseudomonas fluorescens

recombinant esterase detection

AUTHOR: BORNSCHEUER U; BAUMANN M

PATENT ASSIGNEE: BASF AG

PATENT INFO: DE 10124799 28 Nov 2002 APPLICATION INFO: DE 2001-1024799 21 May 2001

PRIORITY INFO: DE 2001-1024799 21 May 2001; DE 2001-1024799 21 May 2001

DOCUMENT TYPE: Patent LANGUAGE: German

OTHER SOURCE: WPI: 2003-343904 [33]

AN 2003-14535 BIOTECHDS AB DERWENT ABSTRACT:

NOVELTY - Detecting hydrolases (I) comprises incubating a sample with an ester (II) of acetic acid with an achiral, chiral or prochiral alcohol, then detecting the acetic acid released.

DETAILED DESCRIPTION - INDEPENDENT CLAIMS are also included for the following: (1) Test kit for the new method comprising, apart from usual components, at least one (pro)chiral substrate (IIa) for (I); and (2) Isolating natural or synthetic hydrolase mutants or variants with altered property profiles.

BIOTECHNOLOGY - Preferred Process: Acetic acid is detected in a coupled enzymatic test, particularly optically. The sample is a crude cell extract, supernatant from a culture of microbial, plant or animal cells, or is derived from a plant, animal, organ or their parts. The entire reaction is done in a microtiter plate. Especially acetic acid is converted enzymatically to acetyl-CoA, which is reacted enzymatically to oxaloacetate to form citrate, where the oxaloacetate is produced from $% \left(1\right) =\left(1\right) \left(1$ L-malate in presence of NAD+ (oxidized nicotinamide-adenine dinucleotide), resulting in formation of reduced NAD (1 mole per mole acetic acid), and this is monitored at 340 nm. (I)-catalyzed formation of acetate is the rate-determining step in the detection process. The method is particularly a high-throughput screen for detecting (I) activity and/or selectivity in extracts of natural or genetically modified organisms, especially to determine enantio- or stereo-selectivity and/or influence of external factors. Preferred Enzymes: (I) is an estearse, lipase, amidase, acylase or protease. Preferred Method: In method (2), a sample is prepared from a prokaryotic or eukaryotic organism and analyzed by the new method. If hydrolase activity is detected, the property profile of the mutant/variant is determined and compared with that for a reference enzyme, and those mutants/variants with altered properties are isolated. The method is particularly applied to recombinant microorganisms that express a hydrolase sequence that has been subjected to

mutagenesis or directed evolution. These are screened for alterations in activity, enantioselectivity, temperature stability and stability in aqueous and/or organic media.

USE - The method is used to detect hydrolases in microbial, plant or animal cells, especially to **isolate** those, produced in recombinant microorganisms by mutagenesis or directed evolution, that have altered properties. The altered enzymes are useful for production of chiral esters and alcohols.

ADVANTAGE - The method can detect variant (I) with improved activity, enantioselectivity and/or stability (to **temperature** or reaction media). It is rapid and inexpensive, especially suitable for high throughput screening of libraries of mutant microorganisms.

EXAMPLE - A recombinant esterase from Pseudomonas fluorescens was tested, in microtiter plates, for hydrolysis of (R,S)alpha-phenylethyl acetate, in presence of acetyl-CoA synthase, citrate synthase, L-malate dehydrogenase, L-malate, adenosine triphosphate, NAD+ (oxidized nicotinamide-adenine dinucleotide) and coenzyme A, to provide a coupled enzymatic system that converts acetate with ultimate formation of citrate, with reduction of NAD+ to NADH. The extinction of NADH at 340 nm was monitored; its rate of change was a linear function of both enzyme concentration and substrate concentration. (13 pages)

L11 ANSWER 11 OF 133 USPATFULL ON STN

ACCESSION NUMBER: 84:44199 USPATFULL

TITLE: Ice nucleating microorganisms INVENTOR(S): Orser, Cindy S., Berkeley, CA

INVENTOR(S):

Orser, Cindy S., Berkeley, CA, United States
Lindow, Steven E., Berkeley, CA, United States
Panopoulos, Nickolas J., Oakland, CA, United States
Stackawicz, Prian L., Castro Valley, CA, United States

Staskawicz, Brian J., Castro Valley, CA, United States
PATENT ASSIGNEE(S): The Regents of the University of California, Berkeley,

CA, United States (U.S. corporation)

APPLICATION INFO.: US 1982-371162
DOCUMENT TYPE: Utility

FILE SEGMENT: Utility
Granted

PRIMARY EXAMINER: Wiseman, Thomas G. ASSISTANT EXAMINER: Martinell, James LEGAL REPRESENTATIVE: Rowland, Bertram I.

NUMBER OF CLAIMS: 13
EXEMPLARY CLAIM: 1
LINE COUNT: 328

PATENT INFORMATION:

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

AB DNA sequences encoding for ice nucleation activity are isolated and introduced into unicellular hosts. The modified hosts demonstrate ice nucleation activity analogous to the DNA source host. The cellular products find use in inhibiting supercooling.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

L11 ANSWER 18 OF 133 CABA COPYRIGHT 2004 CABI on STN

ACCESSION NUMBER:
DOCUMENT NUMBER:

91:61090 CABA 19910505423

TITLE: Alternative

Alternative hosts for Bacillus thuringiensis

delta-endotoxin genes

AUTHOR: Feitelson, J. S.; Quick, T. C.; Gaertner, F.; Baker,

R.R. [EDITOR]; Dunn, P.E. [EDITOR]

CORPORATE SOURCE: Mycogen Corporation, 5451 Oberlin Drive, San Diego,

CA 92121, USA.

SOURCE: UCLA Symposia on Molecular and Cellular Biology,

(1990) Vol. 112, pp. 561-571. 17 ref. Publisher: Alan R. Liss, Inc. New York Price: Conference paper; Journal article

Meeting Info.: New directions in biological control. Alternatives for suppressing agricultural pests and diseases. Proceedings of a UCLA Colloquium held at

19820423 (6)

Frisco, Colorado, January 20-27, 1989.

ISBN: 0-471-56681-0

PUB. COUNTRY: DOCUMENT TYPE: United States

Journal English

LANGUAGE:

Entered STN: 19941101

ENTRY DATE:

Last Updated on STN: 19941101
In general, agricultural application of B. thuringiensis has been limited to the use of formulated spore-crystal mixtures that typically degrade within 1-3 days following application. Degradation appears to be due to a number of factors including: cycles in temperature and humidity, proteolytic and microbial activity, photo-oxidation, and chemical interactions. A novel pesticide delivery system was developed that overcomes these drawbacks by effectively microencapsulating the pesticidal protein within a stabilized Pseudomonas fluorescens cell. Biotoxin genes isolated from B. thuringiensis were introduced into P. fluorescens with the appropriate plasmid vectors. The biotoxin expressed in P. fluorescens formed a crystalline array similar to that seen in B. thuringiensis, with expression levels up to 30%. Unlike B. thuringiensis, the cells of P. fluorescens did not lyse, not did they sporulate, during stationary growth. A chemical fixative was added to the complete fermentation broth to rapidly kill the biotoxin-containing P. fluorescens and to simultaneously stabilize the cells. This stabilization process strengthened the cell wall by crosslinking, and inactivated biotoxin degrading proteolytic enzymes. The process resulted in an active stable biotoxin encapsulated within a nonviable cell. The bioencapsulated products (MCap) exhibited enhanced field persistence and are environmentally acceptable; the microorganism will not spread from the site of application. This delivery system is potentially applicable to a variety of pesticidal proteins.

ANSWER 26 OF 133 BIOTECHDS COPYRIGHT 2004 THOMSON DERWENT/ISI on STN ACCESSION NUMBER: 1995-10495 BIOTECHDS

TITLE:

Extremozymes: expanding the limits of biocatalysis;

thermostable enzyme, psychrophilic enzyme, halophilic enzyme, barophilic enzyme biocatalyst enzyme engineering

and solvent engineering; a review

Adams M W W; Perler F B; *Kelly R M

LOCATION:

CORPORATE SOURCE: Univ.Georgia; New-England-Biolabs; Univ.North-Carolina-State

Department of Chemical Engineering, North Carolina State

SOURCE:

University, Raleigh, NC 27695, USA. Bio/Technology; (1995) 13, 7, 662-68

CODEN: BTCHDA

ISSN: 0733-222X

DOCUMENT TYPE:

Journal English

LANGUAGE: 1995-10495 BIOTECHDS

AR Biocatalysts need not be constrained to mild conditions and can be considered at pH values, temperatures, pressures and in ionic and solvent environments thought to be destructive to biomolecules. It has been shown that even conventional enzymes will catalyze reactions in solvents other than water. The intrinsic basis for biological activity under extreme conditions is only starting to be addressed, as are associated applications. Extremozymes are reviewed with respect to: microorganisms from extreme environments; identification, isolation and production of extremozymes e.g. psychrophilic enzymes, halophilic enzymes, thermostable enzymes and barophilic enzymes; molecular biology of archaea; applications of thermophilic DNA modifying enzymes; cloning and expression of genes encoding extremozymes from thermophilic archaea; mechanisms of extremozyme stability; solvent engineering; high pressure applications; and whole cell biocatalysts. Modification of enzymes to improve their ranges of stability and activity will open new opportunities for using biocatalysis. (86 ref)

L11 ANSWER 28 OF 133 SCISEARCH COPYRIGHT 2004 THOMSON ISI on STN

ACCESSION NUMBER: 95:431831 SCISEARCH

THE GENUINE ARTICLE: RD715

TITLE: ISOLATION OF LUX REPORTER GENE FUSIONS IN

PSEUDOMONAS-FLUORESCENS DF57 INDUCIBLE BY NITROGEN OR

PHOSPHORUS STARVATION

AUTHOR: KRAGELUND L (Reprint); CHRISTOFFERSEN B; NYBROE O; DEBRUIJN F J

CORPORATE SOURCE: MICHIGAN STATE UNIV, NSF CTR MICROBIAL ECOL, E LANSING,

MI, 48824 (Reprint); MICHIGAN STATE UNIV, DEPT ENERGY, PLANT RES LAB, E LANSING, MI, 48824; MICHIGAN STATE UNIV, DEPT MICROBIOL, E LANSING, MI, 48824; ROYAL VET & AGR UNIV, MICROBIOL SECT, DK-1958 FREDERIKSBERG C, DENMARK

COUNTRY OF AUTHOR: USA; DENMARK

SOURCE: FEMS MICROBIOLOGY ECOLOGY, (JUN 1995) Vol. 17, No. 2, pp.

95-106.

ISSN: 0168-6496.
DOCUMENT TYPE: Article; Journal

FILE SEGMENT: LIFE; AGRI LANGUAGE: ENGLISH

REFERENCE COUNT: 39

ABSTRACT IS AVAILABLE IN THE ALL AND IALL FORMATS

We have used transposon Tn5 mutagenesis to insert a promoter-less luxAB gene-cassette into multiple locations in the chromosome of a Pseudomonas fluorescens strain, thereby bringing the Ewe reporter genes under the control of resident promoters. To identify reporter bacteria responsive to nutritional stresses we isolated and characterized a collection of 23 gene fusions consistently displaying bioluminescence under nitrogen starvation and 12 phosphorus starvation inducible fusions. Bioluminescence of one group of mutants was induced after 4 to 6 h of starvation and was continuously expressed at a high level, whereas a second group was induced earlier and the bioluminescence subsequently declined. Finally, a third group was induced later after 24 h of starvation. Four strains were selected for further study, namely, two Tn5-lux containing strains which were induced by nitrogen starvation and two strains induced by phosphorus starvation. Another two strains, carrying constitutively expressed lux fusions, were included as controls. An analysis of biochemical characters, as well as LPS and protein composition, did not reveal any discernible differences between the mutants and the wild-type strain. Survival experiments with the selected Tn5-lux containing strains showed that they all performed comparably to the wild-type under carbon and nitrogen starvation, whereas some of the strains were less resistant to phosphorus starvation. Expression of bioluminescence by the mutants during carbon, nitrogen and phosphorus starvation was detectable

L11 ANSWER 30 OF 133 SCISEARCH COPYRIGHT 2004 THOMSON ISI on STN

ACCESSION NUMBER: 96:408530 SCISEARCH

THE GENUINE ARTICLE: UM108

temperature.

TITLE: CHARACTERIZATION OF MALATE-DEHYDROGENASE FROM DEEP-SEA

even after 18 days and was not affected by high osmolarity or low

PSYCHROPHILIC VIBRIO SP STRAIN NO-5710 AND CLONING OF ITS

GENE

AUTHOR: OHKUMA M (Reprint); OHTOKO K; TAKADA N; HAMAMOTO T; USAMI

R; KUDO T; HORIKOSHI K

CORPORATE SOURCE: INST PHYS & CHEM RES, MICROBIOL LAB, 2-1 HIROSAWA, WAKO,

SAITAMA 35101, JAPAN (Reprint); JAPAN MARINE SCI & TECHNOL CTR, DEEPSTAR PROGRAM, WAKO, SAITAMA 35101, JAPAN; UNIV

TOKYO, DEPT APPL CHEM, KAWAGOE, SAITAMA 350, JAPAN

COUNTRY OF AUTHOR: JAPAN

SOURCE: FEMS MICROBIOLOGY LETTERS, (01 APR 1996) Vol. 137, No.

2-3, pp. 247-252. ISSN: 0378-1097. Article; Journal

DOCUMENT TYPE: Art: FILE SEGMENT: LIFE

LIFE ENGLISH

LANGUAGE: ENGL REFERENCE COUNT: 12

REFERENCE COUNT: 12
ABSTRACT IS AVAILABLE IN THE ALL AND IALL FORMATS

AB A metabolic key enzyme malate dehydrogenase (MDH) was purified from a deep-sea psychrophilic bacterium, Vibrio sp. strain no. 5710. The enzyme displayed an optimal activity shifted toward lower temperature and a pronounced heat lability. A gene encoding this enzyme was isolated and cloned. Recombinant Escherichia coli cells harboring the isolated clone expressed MDH activity with temperature stability identical to that of the parental psychrophile. Nucleotide sequencing of the gene revealed that its primary sequence was

similar to that of a mesophile E. coli MDH (78% amino acid identity), for which the three-dimensional structure is known. The enzyme is thus suitable for the analysis of molecular adaptations to low

L11 ANSWER 34 OF 133 SCISEARCH COPYRIGHT 2004 THOMSON ISI on STN

ACCESSION NUMBER: 97:635383 SCISEARCH

THE GENUINE ARTICLE: XR909

Sequencing and expression of the gene encoding a TITLE:

cold-active citrate synthase from an Antarctic bacterium,

strain DS2-3R

Gerike U; Danson M J; Russell N J; Hough D W (Reprint) AUTHOR:

UNIV BATH, DEPT BIOL & BIOCHEM, CTR EXTRAMOPHILE RES, BATH CORPORATE SOURCE:

BA2 7AY, AVON, ENGLAND (Reprint); UNIV BATH, DEPT BIOL &

BIOCHEM, CTR EXTRAMOPHILE RES, BATH BA2 7AY, AVON,

ENGLAND; UNIV LONDON WYE COLL, DEPT BIOL SCI, ASHFORD TN25

5AH, KENT, ENGLAND

COUNTRY OF AUTHOR:

ENGLAND SOURCE:

EUROPEAN JOURNAL OF BIOCHEMISTRY, (15 AUG 1997) Vol. 248,

No. 1, pp. 49-57.

Publisher: SPRINGER VERLAG, 175 FIFTH AVE, NEW YORK, NY

ISSN: 0014-2956. Article; Journal

DOCUMENT TYPE:

LIFE

FILE SEGMENT: LANGUAGE:

English

29

REFERENCE COUNT:

ABSTRACT IS AVAILABLE IN THE ALL AND IALL FORMATS

The gene encoding citrate synthase from a novel bacterial isolate (DS2-3R) from Antarctica has been cloned, sequenced and over expressed in Escherichia coli. Both the recombinant enzyme and the native enzyme, purified from DS2-3R, are cold-active, with a temperature optimum of 31 degrees C. In addition the enzymes are rapidly inactivated at 45 degrees C, and show significant activity at 10 degrees C and below. Comparison of amino acid sequences indicates that DS2-3R citrate synthase is most closely related to the enzyme from gram-positive bacteria. The amino acid sequence of the DS2-3R enzyme shows several features previously recognised in other cold-active enzymes, including an extended surface loop, an increase in the occurrence of charged residues and a decrease in the number of proline residues in loops. Other changes observed in some psychrophilic enzymes, such as a decrease in isoleucine content and in arginine/(arginine + lysine) content, were not seen in this case.

L11 ANSWER 49 OF 133 SCISEARCH COPYRIGHT 2004 THOMSON ISI on STN

ACCESSION NUMBER: 2000:215771 SCISEARCH

THE GENUINE ARTICLE: 293FE

A bioluminescence assay for screening thermoregulated

genes in a psychrotrophic bacterium Pseudomonas

fluorescens

AUTHOR: CORPORATE SOURCE: Regeard C; Merieau A; GuespinMichel J F (Reprint) FAC SCI ROUEN, LAB MICROBIOL FROID, F-76821 MONT ST

AIGNAN, FRANCE (Reprint); FAC SCI ROUEN, LAB MICROBIOL

FROID, F-76821 MONT ST AIGNAN, FRANCE

COUNTRY OF AUTHOR: FRANCE

SOURCE:

JOURNAL OF APPLIED MICROBIOLOGY, (JAN 2000) Vol. 88, No.

1, pp. 183-189.

Publisher: BLACKWELL SCIENCE LTD, P O BOX 88, OSNEY MEAD,

OXFORD OX2 ONE, OXON, ENGLAND.

ISSN: 1364-5072. Article; Journal

DOCUMENT TYPE: FILE SEGMENT:

LIFE; AGRI

LANGUAGE: REFERENCE COUNT: English 23

ABSTRACT IS AVAILABLE IN THE ALL AND IALL FORMATS AΒ

Random transcription fusion delivery, with bacterial luciferase genes as reporter, was performed in the psychrotrophic bacterium Pseudomonas fluorescens. Direct screening on plates of the insertions allowed the isolation of fusions into

thermoregulated genes with good accuracy, either in a library of

insertion fusions, or after genetic transfer of a

putative regulatory mutation. Using this method, it was shown that in Ps. fluorescens, nearly 40% of the genes are thermoregulated and belong to at least three classes according to the maximal temperature of expression of the fused genes. This is more than had been estimated by a previous method, and demonstrates the importance of thermoregulation in psychrotrophic bacteria. As this reporter is the first to be used for direct screening for genes regulated by temperature, it should be of great value in the study of mechanisms involved in adaptation to this environmental factor.

L11 ANSWER 51 OF 133 SCISEARCH COPYRIGHT 2004 THOMSON ISI on STN

2000:120665 SCISEARCH ACCESSION NUMBER:

THE GENUINE ARTICLE: 281NZ

Cloning of phosphatase I gene from a psychrophile,

Shewanella sp., and some properties of the recombinant

AUTHOR: Tsuruta H; Aizono Y (Reprint)

KOBE UNIV, FAC AGR, DEPT BIOFUNCT CHEM, BIOL CHEM LAB, CORPORATE SOURCE:

NADA KU, KOBE, HYOGO 6578501, JAPAN (Reprint); KOBE UNIV, FAC AGR, DEPT BIOFUNCT CHEM, BIOL CHEM LAB, NADA KU, KOBE,

HYOGO 6578501, JAPAN

COUNTRY OF AUTHOR:

JAPAN

SOURCE: JOURNAL OF BIOCHEMISTRY, (JAN 2000) Vol. 127, No. 1, pp.

143-149.

Publisher: JAPANESE BIOCHEMICAL SOC, ISHIKAWA BLDG-3F,

25-16 HONGO-5-CHOME, BUNKYO-KU, TOKYO 113, JAPAN.

ISSN: 0021-924X. Article; Journal

DOCUMENT TYPE: FILE SEGMENT:

LIFE

LANGUAGE: REFERENCE COUNT: English

ABSTRACT IS AVAILABLE IN THE ALL AND IALL FORMATS

Psychrophilic phosphatase I from Shewanella sp. is a cold enzyme that was found as a novel protein-tyrosine-phosphatase (PTPase, EC 3.1.3.48) with a histidine as its catalytic residue [Tsuruta and Aizono (1999) J. Biochem. 125, 690-695]. Here, we determined the nucleotide sequence of a DNA fragment (2,004 bp) containing the phosphatase I gene by cloning with polymerase chain reaction (PCR) and inverted PCR techniques. The deduced amino acid sequence, of the enzyme contained a conserved region of protein-serine/threonine-phosphatase (PPase). The 38.5 kDa-recombinant protein expressed in Escherichia coli was purified to homogeneity by glutathione-Sepharose 4B column chromatography, treatment with endoproteinase and Mono-Q column chromatography. The recombinant enzyme had a specific activity of 49.4 units and, like native psychrophilic phosphatase I, exhibited high catalytic activity at low temperature and PTPase activity.

L11 ANSWER 65 OF 133 SCISEARCH COPYRIGHT 2004 THOMSON ISI on STN

ACCESSION NUMBER: 2002:924122 SCISEARCH

THE GENUINE ARTICLE: 613CX

TITLE: Characterization of a cloned subtilisin-like serine

proteinase from a psychrotrophic Vibrio species

AUTHOR: Arnorsdottir J; Smaradottir R B; Magnusson O T;

Thorbjarnardottir S H; Eggertsson G; Kristjansson M M

(Reprint)

CORPORATE SOURCE: Univ Iceland, Inst Sci, Dept Biochem, Dunhaga 3, IS-107

Reykjavik, Iceland (Reprint); Univ Iceland, Inst Sci, Dept Biochem, IS-107 Reykjavik, Iceland; Univ Iceland, Inst

Biol, IS-107 Reykjavik, Iceland

COUNTRY OF AUTHOR: Iceland

SOURCE: EUROPEAN JOURNAL OF BIOCHEMISTRY, (NOV 2002) Vol. 269, No.

22, pp. 5536-5546.

Publisher: BLACKWELL PUBLISHING LTD, P O BOX 88, OSNEY

MEAD, OXFORD OX2 ONE, OXON, ENGLAND.

ISSN: 0014-2956.

DOCUMENT TYPE:

Article; Journal

LANGUAGE: REFERENCE COUNT:

English

ABSTRACT IS AVAILABLE IN THE ALL AND IALL FORMATS

AΒ The gene encoding a subtilisin-like serine proteinase in the psychrotrophic Vibrio sp. PA44 has been successfully cloned, sequenced and expressed in Escherichia coli. The gene is 1593 basepairs and encodes a precursor protein of 530 amino acid residues with a calculated molecular mass of 55.7 kDa. The enzyme is isolated, however, a an active 40.6-kDa proteinase, without a 139 amino acid residue N-terminal prosequence. Under mild conditions the enzyme undergoes a further autocatalytic cleavage to give a 29.7-kDa proteinase that retains full enzymatic activity. The deduced amino acid sequence of the enzyme has high homology to proteinases of the proteinase K family of subtilisin-like proteinases. With respect to the enzyme characteristics compared in this study the properties of the wild-type and recombinant proteinases are the same. Sequence analysis revealed that especially with respect to the thermophilic homologues, aqualysin I from Thermus aquaticus and a proteinase from Thermus strain Rt41A, the cold-adapted Vibrio-proteinase has a higher content of polar/uncharged amino acids, a well a aspartate residues. The thermophilic enzymes had a higher content of arginines, and relatively higher number of hydrophobic amino acids and a higher aliphatic index. These factors may contribute to the adaptation of these proteinases to different temperature conditions.

L11 ANSWER 67 OF 133 SCISEARCH COPYRIGHT 2004 THOMSON ISI on STN

ACCESSION NUMBER: 2002:392586 SCISEARCH

THE GENUINE ARTICLE: 547YK

Cloning of cold-active alkaline phosphatase gene of a TITLE:

psychrophile, Shewanella sp., and expression of the

recombinant enzyme

Murakawa T; Yamagata H; Tsuruta H; Aizono Y (Reprint) AUTHOR:

CORPORATE SOURCE: Kobe Univ, Fac Agr, Dept Biofunct Chem, Biol Chem Lab,

Nada Ku, Kobe, Hyogo 6578501, Japan (Reprint)

COUNTRY OF AUTHOR:

SOURCE:

BIOSCIENCE BIOTECHNOLOGY AND BIOCHEMISTRY, (APR 2002) Vol.

66, No. 4, pp. 754-761.

Publisher: JAPAN SOC BIOSCI BIOTECHN AGROCHEM, JAPAN ACAD SOC CTR BLDG, 2-4-6 YAYOI BUNKYO-KU, TOKYO, 113, JAPAN.

ISSN: 0916-8451.

DOCUMENT TYPE:

Article; Journal

LANGUAGE:

English

REFERENCE COUNT:

ABSTRACT IS AVAILABLE IN THE ALL AND IALL FORMATS

A psychrophilic alkaline phosphatase (EC 3.1.3.1) from Shewanella sp. is a cold-active enzyme that has high catalytic activity at low temperature [Ishida et al. (1998) Blosci. Blotechnol. Biochem., 62, 2246-2250]. Here, we identified the nucleotide sequence of a gene encoding the enzyme after cloning with the polymerase chain reaction (PCR) and inverted PCR techniques. The deduced amino acid sequence of the enzyme contained conserved amino acids found among mesophilic alkaline phosphatases and showed some structural characteristics including a high content of hydrophobic amino acid residues and the lack of single alpha-helix compared with the alkaline phosphatase of Escherichia coli, which were possibly efficient for catalytic reaction at low temperatures. The recombinant enzyme expressed in E. coli was purified to homogeneity with the molecular mass of 41 kDa. The recombinant enzyme had a specific activity of 1,500 units/mg and had high catalytic activity at low temperatures.

L11 ANSWER 73 OF 133 USPATFULL on STN

ACCESSION NUMBER: 2002:287601 USPATFULL

TITLE: Enzymes having alpha-galactosidase activity and methods

of use thereof

INVENTOR(S): Short, Jay M., Rancho Santa Fe, CA, UNITED STATES

Murphy, Dennis, Malvern, PA, UNITED STATES Reid, John, Ardmore, PA, UNITED STATES Mathur, Eric J., Carlsbad, CA, UNITED STATES

Diversa Corporation (U.S. corporation) PATENT ASSIGNEE(S):

> NUMBER KIND DATE

PATENT INFORMATION:

US 2002160464

A1 20021031

US 2002-114083 A1 20020401 (10) APPLICATION INFO.:

Division of Ser. No. US 2001-886400, filed on 20 Jun RELATED APPLN. INFO .:

2001, PENDING Continuation-in-part of Ser. No. US 2000-619072, filed on 19 Jul 2000, PENDING Division of Ser. No. US 1999-407806, filed on 28 Sep 1999, PENDING Division of Ser. No. US 1996-613220, filed on 8 Mar

1996, GRANTED, Pat. No. US 5958751

Utility DOCUMENT TYPE:

FILE SEGMENT: APPLICATION

GARY CARY WARE & FRIENDENRICH LLP, 4365 EXECUTIVE LEGAL REPRESENTATIVE:

DRIVE, SUITE 1600, SAN DIEGO, CA, 92121-2189

NUMBER OF CLAIMS:

EXEMPLARY CLAIM:

7 Drawing Page(s) NUMBER OF DRAWINGS:

LINE COUNT: 2958

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

The invention relates to .alpha.-galactosidase and to polynucleotides encoding the .alpha.-galactosidase. In addition methods of designing new .alpha.-qalactosidases and method of use thereof are also provided. The .alpha.-galactosidases have increased activity and stability at

increased pH and temperature.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

ANSWER 99 OF 133 BIOTECHDS COPYRIGHT 2004 THOMSON DERWENT/ISI on STN

ACCESSION NUMBER: 2003-21209 BIOTECHDS

Cloning, heterologous expression, renaturation, and TITLE:

> characterization of a cold-adapted esterase with unique primary structure from a psychrotroph Pseudomonas sp strain

B11-1:

recombinant enzyme production via plasmid expression

SUZUKI T; NAKAYAMA T; CHOO DW; HIRANO Y; KURIHARA T; NISHINO AUTHOR:

T; ESAKI N

CORPORATE SOURCE: Kyoto Univ; Tohoku Univ

Esaki N, Kyoto Univ, Inst Chem Res, Microbial Biochem Lab, LOCATION:

Uji, Kyoto 6110011, Japan

PROTEIN EXPRESSION AND PURIFICATION; (2003) 30, 2, 171-178 SOURCE:

ISSN: 1046-5928

DOCUMENT TYPE: Journal LANGUAGE: English 2003-21209 BIOTECHDS AN

AUTHOR ABSTRACT - A Gene coding for an esterase (PsEst1, 1911 AR bp in length) of the psychrotrophic bacterium Pseudomonas sp. B11-1 isolated from Alaskan soil was cloned and sequenced. The deduced amino acid sequence revealed a protein of $\bar{637}$ amino acid residues with a molecular mass of 69 kDa. Although the expression product, PsEst1, showed no appreciable sequence similarity (less than 15% identity) to any known proteins with the established biochemical functions, it is expected to be related to the alpha/beta hydrolase superfamily because it shared sequence motifs that have been identified with this superfamily. For example, a unique "nucleophilic 6 40 38 elbow"

motif. -Gly(36)-Asp-Ser-Leu-Asn(40)-, was identified, and Ser(38) was predicted to constitute a catalytic triad with Asp(162) and His(303). PsEst1 was overexpressed using a T7 RNA polymerase transcription (pET21a) system in the Escherichia coli BL21(DE3) cells as an inclusion body. A

Soluble denatured form of the enzyme was purified to homogeneity in the presence of 8 M urea, and the catalytically active

form of the enzyme could be obtained by subsequent removal of urea by dialysis, where the addition of 0.1% Triton X-100 was essential for the efficient renaturation of the enzyme. To our knowledge, this was the first example of the successful renaturation of the recombinant cold-adapted enzyme. The enzyme efficiently hydrolyzed vinyl and aryl esters with the C-4-C-6 acyl chain. The activation energy of the enzymatic p-nitrophenyl butyrate hydrolysis (20.1 kcal/mol at 10

degreesC) was significantly lower than the value (79.9 kcal/mol) of the mesophilic lipase. It was observed that the K-m values for p-nitrophenyl

butyrate in the growth temperature range of strain B11-1 (5-15 degreesC) were lower than those at higher temperatures. (C) 2003 Elsevier Science (USA). All rights reserved.

DERWENT ABSTRACT: A 1.9-kbp DNA fragment encoding the PsEstl gene was amplified by polymerase chain reaction (PCR) using

plasmid pUC118-PsEst1 as a template with primers. The entire nucleotide sequence of the amplified DNA was confirmed by sequencing in both orientations. The amplified fragment was then digested with NdeI and BamHI, followed by ligation with NdeI/BamHl-digested pET-21a to produce pET-PsEstl. Escherichia coli BL21 (DE3) cells transformed with pET-PsEstI were cultivated in an Luria-Bertani LB medium containing 200ug/ml ampicillin at 37 deg with shaking. Isopropyl-beta-D-thiogalactopyranoside was added to the medium at a final concentration of 2.0 mM when the turbidity at 600 nm of culture reached 0.8. After another 8 hr cultivation, the cells were harvested. It must be noted that the ${\it expression}$ levels of PsEstl at 15-37 deg did not significantly differ from each other and only inclusion bodies of this protein could be obtained at these temperatures with this hostvector system. Thus, for subsequent renaturation studies, PsLipI was overexpressed at 37 deg, where the host bacterium could grow most rapidly(8 pages)

ANSWER 100 OF 133 BIOTECHDS COPYRIGHT 2004 THOMSON DERWENT/ISI on STN ACCESSION NUMBER: 2003-02464 BIOTECHDS

TITLE:

Cloning and characterization of katA, encoding the major monofunctional catalase from Xanthomonas campestris pv. phaseoli and characterization of theencoded catalase KatA; vector-mediated catalase gene transfer and expression in host cell for recombinant protein production and cloning CHAUVATCHARIN N; VATTANAVIBOON P; SWITALA J; LOEWEN PC;

AUTHOR:

MONGKOLSUK S

CORPORATE SOURCE: Chulabhorn Res Inst; Univ Manitoba; Mahidol Univ

LOCATION:

Mongkolsuk S, Chulabhorn Res Inst, Biotechnol Lab, Lak Si, Bangkok 10210, Thailand

CURRENT MICROBIOLOGY; (2003) 46, 2, 83-87 SOURCE:

ISSN: 0343-8651

DOCUMENT TYPE: Journal LANGUAGE: English 2003-02464 BIOTECHDS AN

AUTHOR ABSTRACT - The first cloning and characterization of the AB gene katA, encoding the majorcatalase (KatA), from Xanthomonas is reported. A reverse genetic approachusing a synthesized katA-specific DNA probe to screen a X. campestris pv. phaseoli genomic library was employed. A positively hybridizing clone designated pKat29 that contained a full-length katA was isolated. Analysis of thenucleotide sequence revealed an open reading frame of 1,521 bp encoding a507-amino acid protein with a theoretical molecular mass of 56 kDa. The deduced amino acid sequence of KatA revealed 84% and 78% identity to CatF of Pseudomonas syringae and KatB of P. aeruginosa, respectively. Phylogenetic analysis places Xanthomonas katA in the clade I group of bacterial catalases. Unexpectedly, expression of katA in a heterologous Escherichia coli hostresulted in a temperature-sensitive expression. The KatA enzyme was purified from an overproducing mutant of X. campestris and was characterized. It has apparent K-m and V-max values of 75 mm [H2O2] and 2.55 X 10(5) mumol H2O2 mumol heme(-1) s(-1), respectively. The enzyme is highly sensitive to 3-amino-1,2,4-triazole

and NaN3, has a narrower optimal pH range than other catalases, and is more sensitive to heat inactivation. (5 pages)

L11 ANSWER 124 OF 133 USPATFULL on STN

ACCESSION NUMBER: 2003:17420 USPATFULL

Novel methods of enzyme purification TITLE:

Gerendash, Joel, San Diego, CA, UNITED STATES INVENTOR(S):

NUMBER KIND DATE US 2003013172 PATENT INFORMATION: A1 20030116 US 2002-146662 APPLICATION INFO.: A1 20020514 (10)

DATE NUMBER

PRIORITY INFORMATION: US 2001-291122P 20010514 (60) DOCUMENT TYPE: Utility

APPLICATION FILE SEGMENT:

LEGAL REPRESENTATIVE: FISH & RICHARDSON, PC, 4350 LA JOLLA VILLAGE DRIVE, SUITE 500, SAN DIEGO, CA, 92122

NUMBER OF CLAIMS:

41

EXEMPLARY CLAIM:

1

NUMBER OF DRAWINGS:

7 Drawing Page(s)

LINE COUNT:

3513

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

The invention relates to alpha amylases and to polynucleotides encoding the alpha amylases. In addition methods of designing new alpha amylases and methods of use and purification thereof are also provided. The alpha amylases have increased activity and stability at increased pH and temperature.

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

=> d his

(FILE 'HOME' ENTERED AT 14:06:56 ON 11 MAR 2004)

INDEX 'ADISCTI, ADISINSIGHT, ADISNEWS, AGRICOLA, ANABSTR, AQUASCI, BIOBUSINESS, BIOCOMMERCE, BIOSIS, BIOTECHABS, BIOTECHDS, BIOTECHNO, CABA, CANCERLIT, CAPLUS, CEABA-VTB, CEN, CIN, CONFSCI, CROPB, CROPU, DISSABS, DDFB, DDFU, DGENE, DRUGB, DRUGMONOG2, ...' ENTERED AT 14:07:12 ON 11 MAR 2004

SEA (PSYCHROTROPH? OR PSYCHROPHIL? OR FLUORESCENS? OR SYRINGAE?

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129*
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      FILE SCISEARCH
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FILE TOXCENTER FILE USPATFULL

FILE USPAT2 FILE VETU

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QUE (PSYCHROTROPH? OR PSYCHROPHIL? OR FLUORESCENS? OR SYRINGAE? L1FILE 'SCISEARCH, CABA, CAPLUS, BIOTECHNO, LIFESCI, ESBIOBASE, GENBANK, DGENE, BIOSIS, EMBASE, USPATFULL, PASCAL, BIOTECHDS, AGRICOLA, MEDLINE' ENTERED AT 14:11:33 ON 11 MAR 2004 9026 S (PSYCHROTROPH? OR PSYCHROPHIL? OR FLUORESCENS? OR SYRINGAE?) L22256 S L2 (S) (ISOLAT? OR PURIF?) L3 1886 S L3 (S) (VECTOR? OR GENE? OR POLYNUCLEOT? OR INSERT? OR DNA? L4732 S L4 (S) FLUORESCENS? L5 368 S L5 (S) COLI L6 161 DUP REM L6 (207 DUPLICATES REMOVED) L7 103 S L7 AND PY<=1999 $^{\text{L8}}$ 259 S L4 (S) (HEAT? OR TEMPERAT?) L9 133 DUP REM L9 (126 DUPLICATES REMOVED) L10 133 SORT L10 PY A L11 => log h COST IN U.S. DOLLARS SINCE FILE TOTAL ENTRY SESSION 236.88 FULL ESTIMATED COST 232.68

SESSION WILL BE HELD FOR 60 MINUTES
STN INTERNATIONAL SESSION SUSPENDED AT 15:11:59 ON 11 MAR 2004

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FILE WPINDEX FILE NLDB

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<u>L7</u>	11 and nano.in.	1	<u>L7</u>
<u>L6</u>	L2 same (heat\$3 or temperatur\$3)	69	<u>L6</u>
<u>L5</u>	L2 same (fLUORESCENs or SYRINGAE) same (heat\$3 or temperatur\$3)	14	<u>L5</u>
<u>L4</u>	L2 same (fLUORESCENs or SYRINGAE)	113	<u>L4</u>
<u>L3</u>	L2 same (fLUORESCENS\$3 or SYRINGAE\$4)	123	<u>L3</u>
<u>L2</u>	L1 same (vector\$3 or gene\$3 or polynucleot\$4 or insert\$3 or dna\$3 or plasmid\$3)	204	<u>L2</u>
<u>L1</u>	(PSYCHROTROPH\$4 OR PSYCHROPHIL\$4 OR FLUORESCENS\$4 OR SYRINGAE\$4) same EXPRESS\$4 same (ISOLAT\$4 OR PURIF\$4)	233	<u>L1</u>

END OF SEARCH HISTORY

DATE: Thursday, March 11, 2004 Printable Copy Create Case

Freeform Search

Search History

Clear Interrupt.

DATE: Thursday, March 11, 2004 Printable Copy Create Case

Search

Set Name side by side	Query	<u>Hit</u> <u>Count</u>	Set Name result set
DB = 1	DWPI; PLUR=YES; OP=OR		
<u>L25</u>	9900492	10	<u>L25</u>
DB =	USPT; PLUR=YES; OP=OR		
<u>L24</u>	(5459055 or 5786174 or 5872238 or 5969121 or 5981177).pn.	5	<u>L24</u>
<u>L23</u>	6294358.pn.	1	<u>L23</u>
<u>L22</u>	L16 and (promote\$4 and (pseudomon\$3 or coli or fluorescens or aeruginosa or syringae or putida)).ti.	0	<u>L22</u>
<u>L21</u>	L18 and (promote\$4 and (pseudomon\$3 or coli or fluorescens or aeruginosa or syringae or putida)).ti.	0	<u>L21</u>
<u>L20</u>	L18 and (promote\$4 and pseudomon\$3 or coli or fluorescens or aeruginosa or syringae or putida).ti.	8	<u>L20</u>
<u>L19</u>	L18 and (promote\$4).ti.	134	<u>L19</u>
<u>L18</u>	(method\$3 or proces\$4) same (promote\$4 or promoto\$4) same (screen\$4 or isolat\$4 or identif\$5) same (reporte\$3 or (select\$4 same marker\$4))	2142	<u>L18</u>
<u>L17</u>	L16.ti.	0	<u>L17</u>
<u>L16</u>	(method\$3 or proces\$4) same (promote\$4 or promoto\$4) same (screen\$4 or isolat\$4) same (reporte\$3 or (select\$4 same marker\$4))	1757	<u>L16</u>

<u>L15</u>	L14 same (identif\$4)	71	<u>L15</u>
<u>L14</u>	L12 same (method\$ or proce\$4)	352	<u>L14</u>
<u>L13</u>	L12 same (pseudomon\$3 or coli or fluorescens or aeruginosa or syringae or putida)	138	<u>L13</u>
<u>L12</u>	L11	1027	<u>L12</u>
DB =	USPT,EPAB,DWPI; PLUR=YES; OP=OR		
<u>L11</u>	L10 same (luciferas\$3 or galactosidas\$4 or gfp\$3)	1055	<u>L11</u>
<u>L10</u>	L9 same (isolat\$4 or clon\$3 or characteri\$5)	8128	<u>L10</u>
<u>L9</u>	promot\$3 same (reporte\$4 or (select\$4 same marke\$4))	19729	<u>L9</u>
<u>L8</u>	L7 same (contamin\$4 or impurit\$4 or undesire\$3)	1	<u>L8</u>
<u>L7</u>	L6 same (cell\$3 or organism\$3 or host\$3)	208	<u>L7</u>
<u>L6</u>	taq\$2 same polymeras\$3 same (heat\$3 or temperatur\$4) same (inactivat\$3 or denatur\$4)	947	<u>L6</u>
DB =	EPAB; PLUR=YES; OP=OR		
<u>L5</u>	373962	1	<u>L5</u>
DB =	USPT; PLUR=YES; OP=OR		
<u>L4</u>	L2 heat\$3 same inactiv\$4 same protei\$3 same (cell\$3 or organism\$3)	1299	<u>L4</u>
<u>L3</u>	(fluorescens or aeruginosa or syringae or putida) same heat\$3 same (inactiv\$4 or denatur\$3)	30	<u>L3</u>
<u>L2</u>	heat\$3 same inactiv\$4 same protei\$3 same cell\$	1266	<u>L2</u>
L1	6080564.pn.	1	<u>L1</u>

END OF SEARCH HISTORY

Hit List

Clear Generate Collection , Print Fwd Refs Bkwd Refs
Generate OACS

Search Results - Record(s) 65 through 74 of 113 returned.

☐ 65. Document ID: US 5952208 A

Using default format because multiple data bases are involved.

L4: Entry 65 of 113

File: USPT

Sep 14, 1999

US-PAT-NO: 5952208

DOCUMENT-IDENTIFIER: US 5952208 A

TITLE: Dsz gene expression in pseudomonas hosts

DATE-ISSUED: September 14, 1999

INVENTOR-INFORMATION:

CITY ZIP CODE COUNTRY NAME STATE Darzins; Aldis The Woodlands ΤX Xi; Lei The Woodlands TXChilds; John D. The Woodlands TXThe Woodlands Monticello; Daniel J. TXSquires; Charles H. The Woodlands TX

US-CL-CURRENT: 435/156; 435/252.34, 435/282

Full	Title	Citation	Front	Review	Classification	Date	Reference	Sequerioss	vatraduname	Claims	KWMC	Drawt De
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☐ 66. Document ID: US 5939601 A

L4: Entry 66 of 113

File: USPT

Aug 17, 1999

US-PAT-NO: 5939601

DOCUMENT-IDENTIFIER: US 5939601 A

TITLE: Genes associates with enhanced disease resistance in plants

DATE-ISSUED: August 17, 1999

INVENTOR-INFORMATION:

NAME CITY

STATE ZIP CODE

COUNTRY

Klessig; Daniel F.

Bridgewater

NJ

Yang; Yinong

Piscataway

NJ

US-CL-CURRENT: 800/279; 435/252.2, 435/320.1, 435/469, 435/470, 536/23.6

Full Title Citation Front Review Classification Date Reference Scribences Attachnicities Claims KWIC Draw, De

☐ 67. Document ID: US 5932698 A

L4: Entry 67 of 113

File: USPT

Aug 3, 1999

US-PAT-NO: 5932698

DOCUMENT-IDENTIFIER: US 5932698 A

TITLE: Recombinant gene coding for a protein having endochitinase activity

DATE-ISSUED: August 3, 1999

INVENTOR-INFORMATION:

STATE ZIP CODE COUNTRY NAME CITY FR Dubois; Michel Buc Escalquens FR Grison; Rene Auzeville Tolosane FR Leguay; Jean-Jacques FR Pignard; Annie Roquettes FR Cornebarrieu Toppan; Alain

US-CL-CURRENT: $\underline{530}/\underline{350}$; $\underline{435}/\underline{200}$, $\underline{435}/\underline{201}$, $\underline{435}/\underline{418}$, $\underline{435}/\underline{419}$, $\underline{435}/\underline{69.1}$, $\underline{435}/\underline{69.7}$, 530/370, 530/379, 536/23.4, 536/23.6

Full Title	Citation Front	Review Classification	Date Reference	Bengendes Altachmen	65 Claims KW	MC Dravu De
Г 68	Document ID	: US 5932209 A				
	68 of 113	. 03 3732207 A	File:	USPT	Aug 3,	. 1999

US-PAT-NO: 5932209

DOCUMENT-IDENTIFIER: US 5932209 A

** See image for Certificate of Correction **

TITLE: Bacillus thuringiensis .delta.-endotoxin

DATE-ISSUED: August.3, 1999

INVENTOR-INFORMATION:

ZIP CODE COUNTRY CITY STATE NAME Del Mar CA Thompson; Mark Schwab; George E. La Jolla CA Schnepf; H. Ernest San Diego CA Stockhoff; Brian San Diego CA

US-CL-CURRENT: 424/93.2; 424/832, 424/93.4, 424/93.461, 435/252.3, 514/12, 530/350, 530/825

Jan 12, 1999

Full Title Citation Front Review Classification Date Reference Sequences Attachments Claims KWC Draw, De

File: USPT

US-PAT-NO: 5859340

L4: Entry 69 of 113

DOCUMENT-IDENTIFIER: US 5859340 A

TITLE: Recombinant gene coding for a protein having endochitinase activity

DATE-ISSUED: January 12, 1999

INVENTOR-INFORMATION:

STATE ZIP CODE COUNTRY CITY NAME FR Dubois; Michel Buc Grison; Rene Escalquens FR FR Leguay; Jean-Jacques Auzeville Tolosane FR Pignard; Annie Roquettes FR Toppan; Alain Cornebarrieu

US-CL-CURRENT: 800/279; 435/200, 435/414, 435/416, 435/418, 435/419, 435/69.1, 435/69.7, 435/69.8, 435/70.1, 536/23.2, 536/23.4, 536/23.6, 536/24.1, 800/301

	Full	Title	Citation	Front	Review	Classification	Date	Reference	Sepudices Alechindric	Claims KV	MC Draw, De
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☐ 70. Document ID: US 5858786 A

L4: Entry 70 of 113

File: USPT

Jan 12, 1999

US-PAT-NO: 5858786

DOCUMENT-IDENTIFIER: US 5858786 A

TITLE: Pseudomonas syringae pv Syrinagae hrpZ gene

DATE-ISSUED: January 12, 1999

INVENTOR-INFORMATION:

NAME CITY STATE ZIP CODE COUNTRY

Collmer; Alan Ithaca NY He; Sheng-Yang Lexington KY

US-CL-CURRENT: 800/298; 435/252.3, 435/320.1, 435/325, 435/418, 435/69.1, 435/71.2, 435/874, 536/23.1, 536/23.7, 800/301

Full Title Citation Front Review Classification Date Reference Sequences Attachnicots Claims KVMC Draw. De

71. Document ID: US 5840554 A

Page 4 of 5

L4: Entry 71 of 113

File: USPT

Nov 24, 1998

US-PAT-NO: 5840554

DOCUMENT-IDENTIFIER: US 5840554 A

** See image for Certificate of Correction **

TITLE: .beta.-Endotoxin expression in pseudomonas fluorescens

DATE-ISSUED: November 24, 1998

INVENTOR-INFORMATION:

NAME

CITY

STATE ZIP CODE

COUNTRY

Thompson; Mark

Del Mar

CA

Schwab; George E.

La Jolla

CA

US-CL-CURRENT: $\underline{435}/\underline{471}$; $\underline{424}/\underline{405}$, $\underline{424}/\underline{538}$, $\underline{435}/\underline{252.34}$, $\underline{435}/\underline{320.1}$, $\underline{435}/\underline{480}$, $\underline{435}/\underline{69.7}$, 514/2, 530/350, 536/23.4, 536/23.71

Full	Title	Citation Front F	Review Classificat	ion Date	Reference	86419 (N-S)	. Site shines to	Claims	KWIC	Drawii De
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				······································						
	72.	Document ID:	US 5827514	A						

L4: Entry 72 of 113

File: USPT

Oct 27, 1998

US-PAT-NO: 5827514

DOCUMENT-IDENTIFIER: US 5827514 A

** See image for Certificate of Correction **

TITLE: Pesticidal compositions

DATE-ISSUED: October 27, 1998

INVENTOR-INFORMATION:

NAME

Schwab; George E.

CITY

STATE ZIP CODE COUNTRY

Bradfisch; Gregory A.

San Diego

CA CA

Thompson; Mark

San Diego La Jolla

CA

US-CL-CURRENT: 424/93.2; 424/93.1, 424/93.3, 435/252.3, 435/410, 435/418, 435/419, <u>435/69.1</u>, <u>435/69.7</u>

Full Title Citation Front Review Classification Date Reference Segmentars Attachinism	Claims	KWIC	Draw, De

☐ 73. Document ID: US 5817502 A

L4: Entry 73 of 113

File: USPT

Oct 6, 1998

US-PAT-NO: 5817502

DOCUMENT-IDENTIFIER: US 5817502 A

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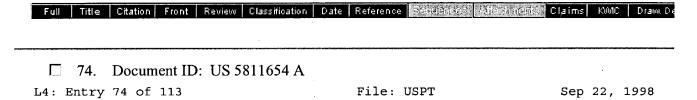
TITLE: Genes for the synthesis of pyrrolnitrin

DATE-ISSUED: October 6, 1998

INVENTOR-INFORMATION:

CITY STATE ZIP CODE COUNTRY NAME NC Ligon; James M. Apex Hill; Dwight Steven Cary NC NC Lam; Stephen Ting Raleigh Hammer; Philip E. Cary NC van Pee; Karl-Heinz Bannewitz DEDE Kirner; Sabine Puchheim

US-CL-CURRENT: 435/252.34; 435/117, 435/252.3, 435/252.33, 435/320.1, 435/69.1, 435/71.1, 536/23.2, 536/23.7



US-PAT-NO: 5811654

DOCUMENT-IDENTIFIER: US 5811654 A

TITLE: Plants genetically enhanced for nutritional quality

DATE-ISSUED: September 22, 1998

INVENTOR-INFORMATION:

NAME

CITY

STATE

COUNTRY

ZIP CODE

Jaynes; Jesse M.

Baton Rouge

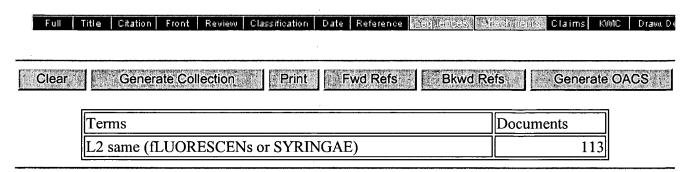
LA

Derrick; Kenneth S.

Lake Alfred

ਜ਼ਾ

US-CL-CURRENT: 800/298; 435/419, 435/69.1, 800/301



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